

<b>ACGT MICROARRAY FACILITY</b>		
<b>STANDARD OPERATING PROCEDURE</b>		
<b>TITLE: cDNA Synthesis and labelling</b>		<b>PAGE: 1 of 2</b>
<b>SOP#: MA005</b>	<b>REVISION LEVEL: .1</b>	<b>DATE: 12 December 2005</b>
<b>AUTHOR: Danielle Tiedt</b>	<b>REVIEWERS: Luke Solomon, Erica Pierce</b>	

### 1. PURPOSE

This procedure describes RNA isolation from human cell lines.

### 2. SCOPE

This protocol has been modified in order to obtain high quality RNA without proteins, which clog the membranes of the clean-up columns.

### 3. MATERIALS

Qiazol/ TriReagent (Qiagen Scientific, catalogue number 79306 or Sigma; cat# T9424)  
RNeasy RNA isolation Kit (Qiagen; cat#74104)

### 4. PREPARATION

### 5. PROCEDURE

#### Harvest Cells

Remove Medium  
Wash cells 2x with 5ml PBS

#### Disrupt Cells

Add 1ml RLT buffer to flask (adding  $\beta$ -ME is preferable)  
Decant flask until all cells have lysed

#### Homogenize Cells

Place QIAshredder into 2ml collection tube  
Pipette lysate into QIAshredder  
Centrifuge for 2min @ max speed (12000g)  
Pool flow through into large 50ml (Falcon) tube

#### Triagent Step

Add 1ml Triagent/Qiazol for each collection tube used in step 3  
Store for 5min at room temperature  
Add 0.3ml chloroform for each collection tube used in step 3  
Cover the sample and shake vigorously  
Store @ room temperature for 2-15min

Centrifuge for 15 min @ 4°C @12000g  
Remove the upper aqueous phase into a 50ml or 10ml tube

<b>ACGT MICROARRAY FACILITY</b>		
<b>STANDARD OPERATING PROCEDURE</b>		
<b>TITLE: cDNA Synthesis and labelling</b>	<b>PAGE: 2 of 2</b>	
<b>SOP#: MA005</b>	<b>REVISION LEVEL: .1</b>	<b>DATE: 12 December 2005</b>
<b>AUTHOR: Danielle Tiedt</b>	<b>REVIEWERS: Luke Solomon, Erica Pierce</b>	

#### Precipitation

Add 1 Volume 70% ethanol  
Mix well (Don't Centiguge)  
Apply the mixture to the Qiagen columns (700µl per column)  
Centrifuge for 15sec @ 10900rpm  
Discard flow through

#### Wash Step 1

Add 350µl RW1 buffer to each column  
Centrifuge for 15sec @ 10900rpm  
Add DNase1 Mixture (70µl RDD buffer + 10µl DNase1)  
NB!! Add to middle of column!!!  
Store @ room Temp for 15min  
Add another 350µl RW1 buffer to each column  
Centrifuge for 15sec A10900rpm  
Transfer to new sterile collection tubes

#### Wash Step 2

Add 500µl of RPE Buffer to each column  
Centrifuge for 15sec @ 10900rpm  
Discard the flow through  
Transfer columns to new sterile collection tubes

#### Wash Step 3

Add another 500µl RPE buffer to each column  
Centrifuge for 15sec @ 10900rpm  
Transfer columns to new sterile collection tubes

#### Dry Step

Centifuge for 1min at full speed  
Transfer the columns to new sterile eppies

#### Elution of RNA

Add 50 $\mu$ l of RNase- free water directly onto each column

Centrifuge for 1min at 10900rpm

The flow- through liquid contains the RNA

Repeat step to increase yield